1. Whole water, filtered water and filters for each site:
   * 3 GF/F filters in labeled foil packets (store in -20˚C freezer)
   * 2 x 40 ml GF/F filtrate for nutrients (store in -20˚C freezer)
   * 2 x 40 ml whole water for nutrients + in-house toxin analysis (store in -20˚C freezer)
   * 1 x 40 ml whole water for Loftin USGS lab. MUST BE STORED IN TUBE THAT HAS BEEN TRIPLE RINSED WITH SAMPLE WATER. (store in -20˚C freezer)
   * 1 x 40 ml DI water for Loftin USGS lab in TRIPLE RINSED tube.
   * 2 G/FF filters in labeled microcentrifuge tubes for DNA (store in -80˚C freezer)
   * 1 0.2 µm Pall filter for DNA (store in -80˚C freezer).
2. Preserve phytoplankton
   * Lugol’s for microscopy
     + 10 ml whole water in 15 ml opaque tube with 100 µl lugol’s solution added
3. Read water on Turner fluorometer and record on fluorometry record sheet. First let water come to room temperature to avoid fogging on cuvette.
   * Turbidity
   * Chl-a (in vivo)
4. Transfer zooplankton samples to scintillation vials.
   * DNA sample: For one of the two replicate zooplankton samples from each site, filter the zooplankton sample to remove as much liquid as possible and re-preserve in 95% ethanol in scintillation vial.
   * Morphology sample: For the second replicate, filter the zooplankton sample and re-preserve in 80% ethanol + 1 % glyceral in scientillation vial.
   * Record site, date, tow depth and number of tows, and net size on two small pieces of card stock in pencil and add to the scintillation vials. Label vial top with site, date, and sample type (DNA or morph).